



LACTATE DETECTION USING MICROWAVE SPECTROSCOPY FOR IN-SITU MEDICAL APPLICATIONS

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Submitted: June 30, 2011

Accepted: July 30, 2011

Published: September 1, 2011

Abstract –Lactate is an indicator of neurological impairment during aortic aneurysm surgery, and thus its detection could provide the basis for improved treatment regimes, better quality of care, and enhanced resource efficiency within the UK National Health Service (NHS). This article considers the use of low power microwave sensors to detect varying lactate concentrations in distilled water; sample sizes of 10ml and 0.5ml are considered via two sensor designs. Microwave sensors provide a rapid non-invasive method of material analysis, which is robust and cost-effective, in addition to harbouring huge potential for a wide range of biomedical applications.

Index terms: microwave, cavity, sensor, lactate, cerebrospinal fluid.

surgeons information upon which they may act quickly. This has the potential to reduce further invasive procedures for that patient, whilst increasing hospital efficiency, allowing them to serve other patients.

Microwave analysis (or microwave spectroscopy) has a range of advantages for biomedical applications. It is a non-ionising technique utilising low power output at around 1mW (0dBm) but has good penetration depth and equipment can be portable for use at the bedside. The multi-parameter nature of broadband microwave analysis can provide unique signal spectrum signatures which are a reflected signal, $|S_{11}|$, and/or a transmitted signal, $|S_{21}|$, based on parameters such as conductivity and permittivity [8]. Conductivity is a measurement of a material's ability to conduct an electric current. Permittivity is a measurement of how an electric field is affected by a dielectric medium, which is determined by the ability of a material to polarise in response to the field, and reduce the total electric field inside the material. Therefore, permittivity relates to a material's ability to transmit an electric field and is a complex value which varies with changing frequency, and accounts for both the energy stored by a material (ϵ') as well as any losses of energy (ϵ'') which might occur.

As a precursor to testing on human samples, this work looks at the microwave response of varying levels of lactate in water, to determine whether it can indeed be used as a marker of ischemia in CSF. This is partly due to lactate being inexpensive, in addition to CSF providing a relative simple background which provides a good basis for working with human samples.

The remaining parts of this paper consider the theory of operation underpinning the work, the experimental procedures undertaken and also the results obtained from the work.

II. MICROWAVE THEORY AND OPERATION

As a material alters in concentration or type, it is likely that its permittivity will change. This leads to a change in response if the material is the target of microwave radiation. By measuring this change in response over a range of frequencies, one can characterise materials in order to infer their properties. Figure 1 depicts the basic principles of a microwave cavity, showing the

permittivity will result in a change in resonant frequency. This is due to the reliance of TE and TM modes on $\sqrt{\epsilon_r}$, and therefore results in resonant peaks shifting to lower frequencies as permittivity increases.

$$f_{nml} = \frac{c}{2\pi\sqrt{\mu_r\epsilon_r}} \sqrt{\left[\left(\frac{p_{nm}}{a}\right)^2 + \left(\frac{l\pi}{d}\right)^2\right]} \quad (1)$$

where:

c is the speed of light

μ_r is of the relative permeability

ϵ_r is the relative permittivity

p_{nm} is the value of the Bessel function for the TE or TM modes of a circular waveguide

a is the radius of the cavity

d is the height of the cavity

III. EXPERIMENTAL METHODOLOGY

a. Microwave Cavities

Two microwave cavities have been designed and used for this research. These are shown in Figure 2; (a) shows a cavity designed to accept large volume samples, and (b) shows a cavity designed to accept smaller volumes. Ultimately the latter is preferential in medical situations since the availability of many biological materials is minimal; CSF however is an exception to this rule, with 10-15ml being readily available from patients as a result of the draining procedure which takes place during surgical or endovascular aneurysm repair at LHCH. The samples are discussed further later in the paper.

The large volume cavity is capable of analysing samples of up to 15ml in volume – smaller sample sizes are possible, however since the sample receptacle is fully immersed in the cavity the sample volumes must be identical for reliable comparison. This cavity has two ports, therefore allowing simple measurement of the $|S_{11}|$ and $|S_{21}|$ spectra.

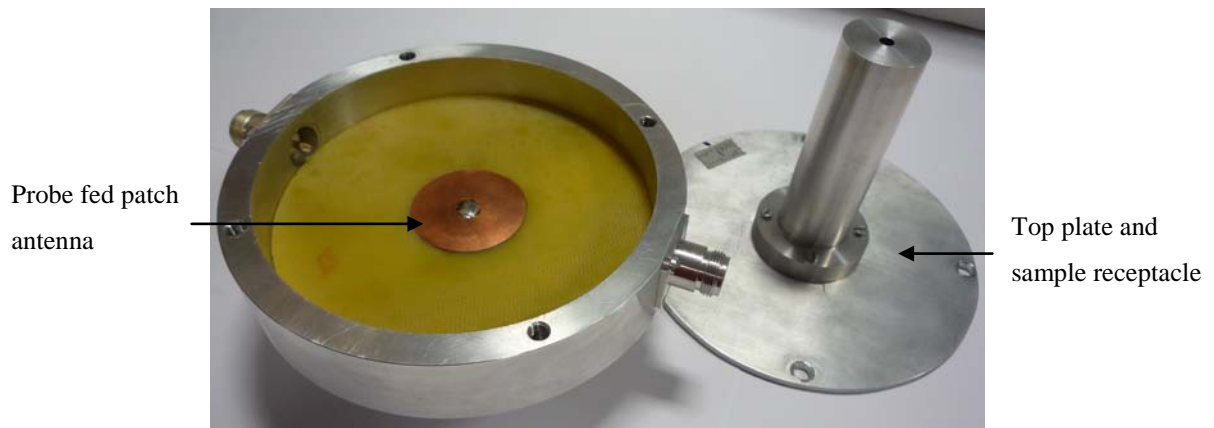
The smaller volume cavity can accept sample receptacles of up to 2.3ml in volume, however the internal height of the cavity means that practically only the lowest 0.5ml of that volume is



(a)



(b)



(c)

Figure 2: Two cavities have been used during this research work in order to test a different volume of sample, (a) large volume cavity, (b) small volume cavity. The innards of the small volume cavity are shown in (c).

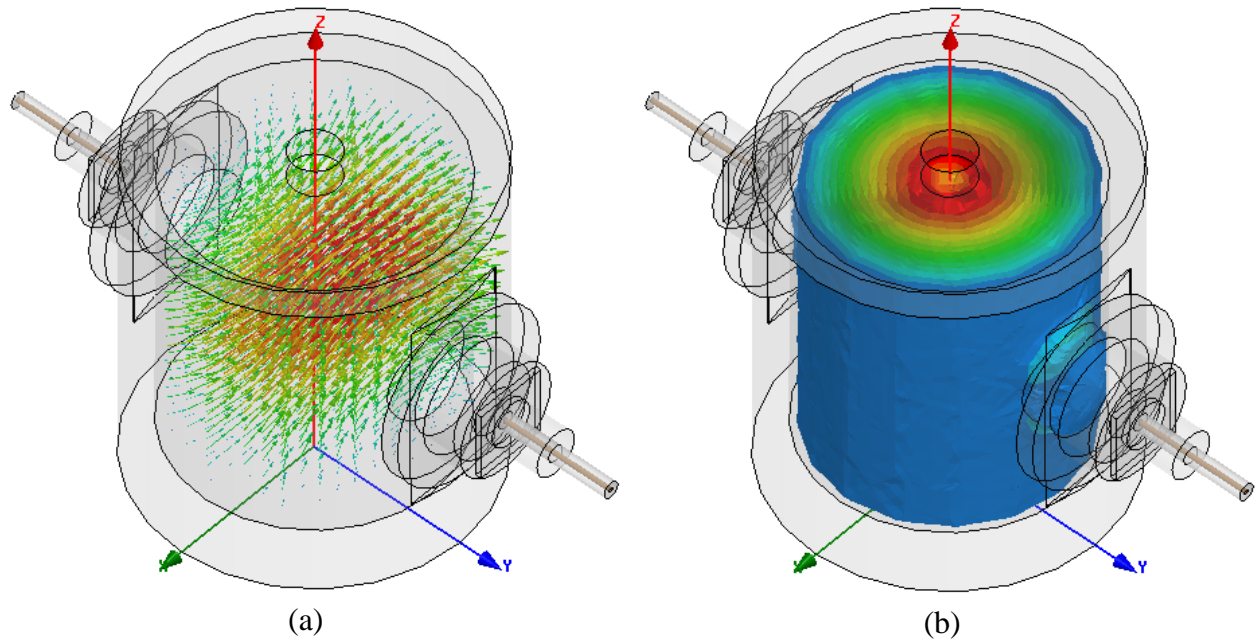


Figure 3: TE and TM modes in the empty large volume cavity, (a) TE_{111} , (b) TM_{010}

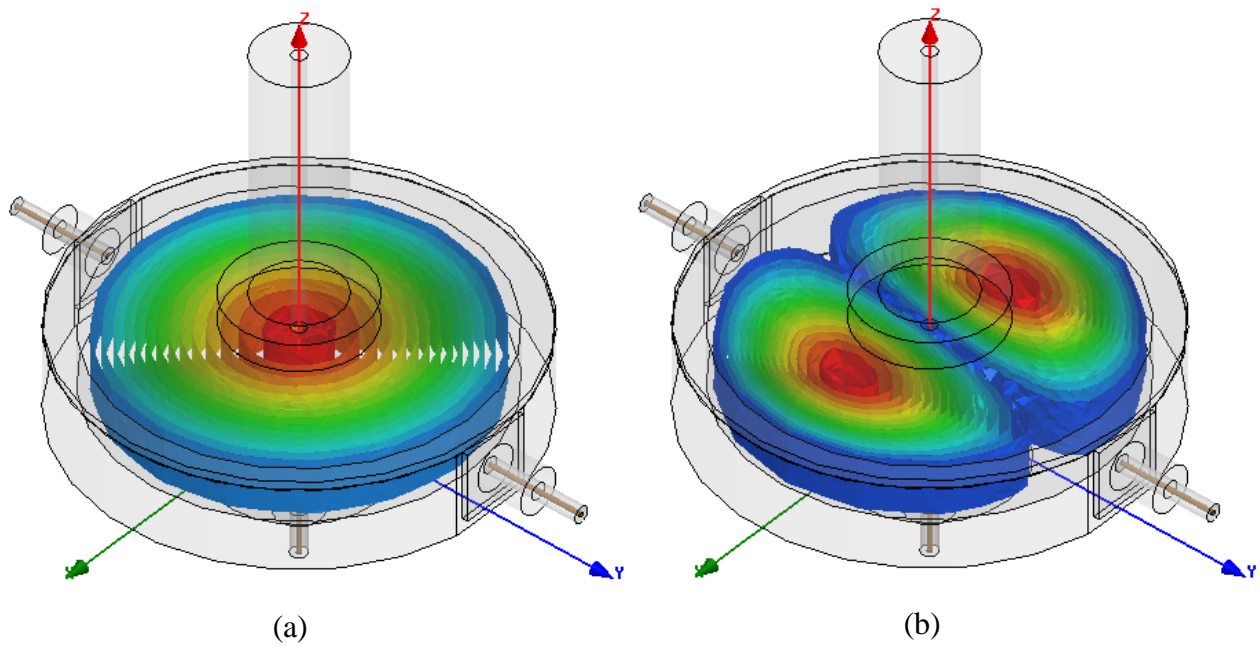


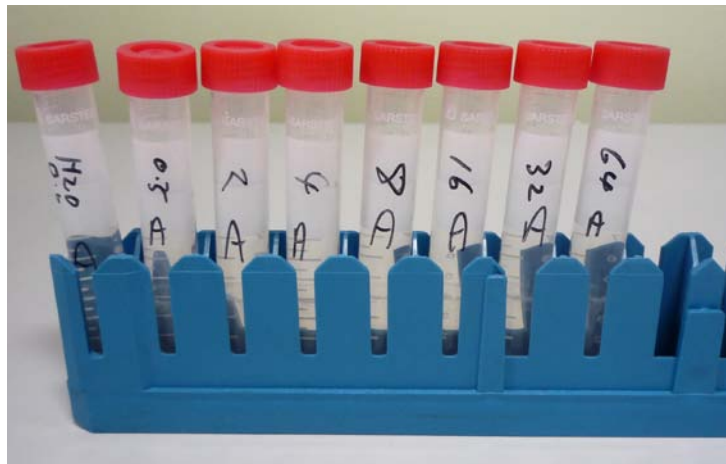
Figure 4: TM modes in the empty small volume cavity, (a) TM_{010} , (b) TM_{110}

b. Lactate Preparation

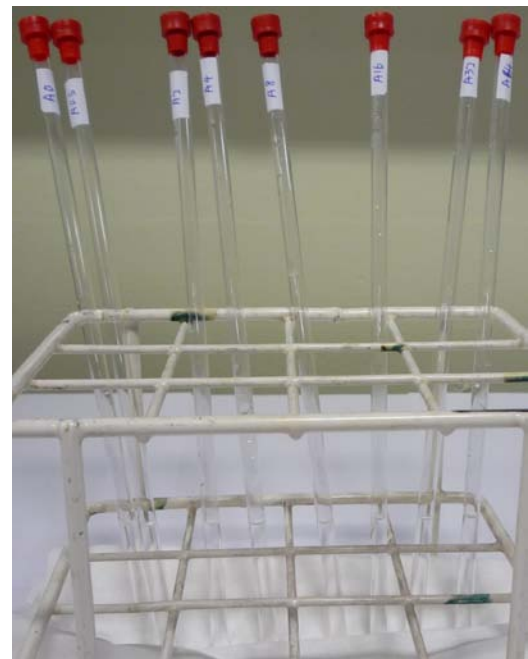
LHCH has taken responsibility for preparing all of the samples used in this work, and they send them to LJMU laboratories for analysis. Serial dilutions of L(+)-lactic acid were prepared to cover a range from low, physiological and supra-physiological levels (0-64mM) in distilled water. Sample concentrations were as follows: 0mM (distilled water), 0.5mM, 2mM, 4mM, 8mM, 16mM, 32mM, 64mM and 1M.

The samples were stored in both 15 ml polypropylene centrifuge tubes and 2.3ml glass tubes, as shown in Figure 5(a) and 5(b) respectively. The former contained 10ml of fluid, whilst the later contained 0.5ml.

All of the samples were stored in a refrigerated unit at 5°C. Samples were removed only to be exposed to the cavity for a short period of time, after which they were returned to the refrigerator. Prior to testing, samples were inspected briefly for spoiling (e.g. due to bacterial growth).



(a)



(b)

Figure 5: Shows the lactate samples provided by LHCH in (a) 15ml polypropylene centrifuge tubes, and (b) 2.3ml glass tubes, utilised to fit the large and small volume cavities respectively.

c. Equipment Setup

All measurements were performed using a ZVL-6 Rohde and Schwarz Vector Network Analyser (VNA). The instrument can generate and sample frequencies up to 6GHz. The full spectrum was swept for each sample, although results shown later are related to areas within the spectrum which are of interest. Figure 6 shows the experimental setup for lactate measurement using the large volume cavity; the only difference between this setup and that of the small volume cavity is the single coaxial connection (instead of dual connection, as shown in Figure 6) between the cavity and VNA. Temperature variations were minimised through the use of an environmental chamber set to an arbitrary constant temperature of 20°C.

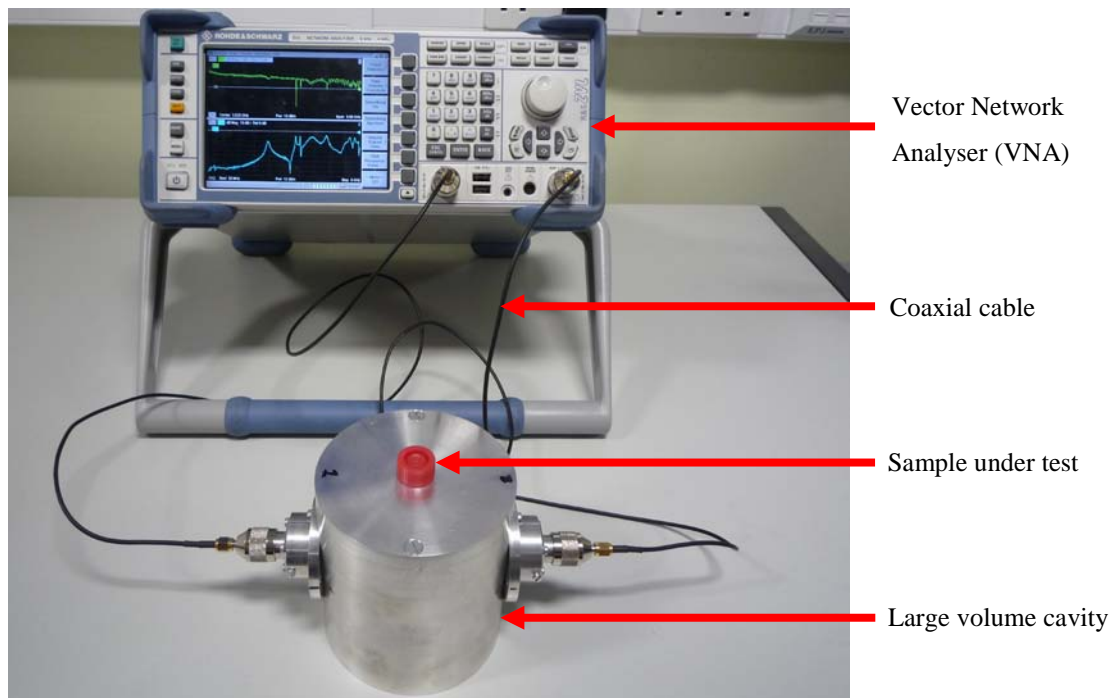


Figure 6: A sample of lactate in water analysed using the large volume cavity, showing the major components of the experimental setup.

IV. EXPERIMENTAL RESULTS

a. Large volume cavity

Figure 7 shows the $|S_{11}|$ spectrum for the large volume cavity, which was used to analyse the 10ml samples. From this spectrum, one can see the level of amplitude decrease as the concentration of the lactate increases from 0mM (distilled water) to 64mM at 3.461GHz. These

results demonstrate a linear sensor response to varying concentrations of lactate in distilled water. The highest amplitude (-27.31dB) occurs when analysing the 0mM sample, whilst the lowest occurs for the 64mM sample (-29.85dB). Across the complete sample set, the amplitude decreases with lactate concentration at a rate of 0.04dB/mM.

The mode at which this response occurs can be determined by matching the measured resonance to that of the simulated model in HFSS. Unlike in section III however, it is necessary to include the sample container with 10ml water in the model in order to determine the correct mode. Figure 8 shows the simulated model for the large volume cavity at 3.458GHz, where the TE_{211} mode dominates. Whilst there is 3MHz difference between simulated and measured resonance, this is considered to be a tolerable error which does not impact on mode determination.

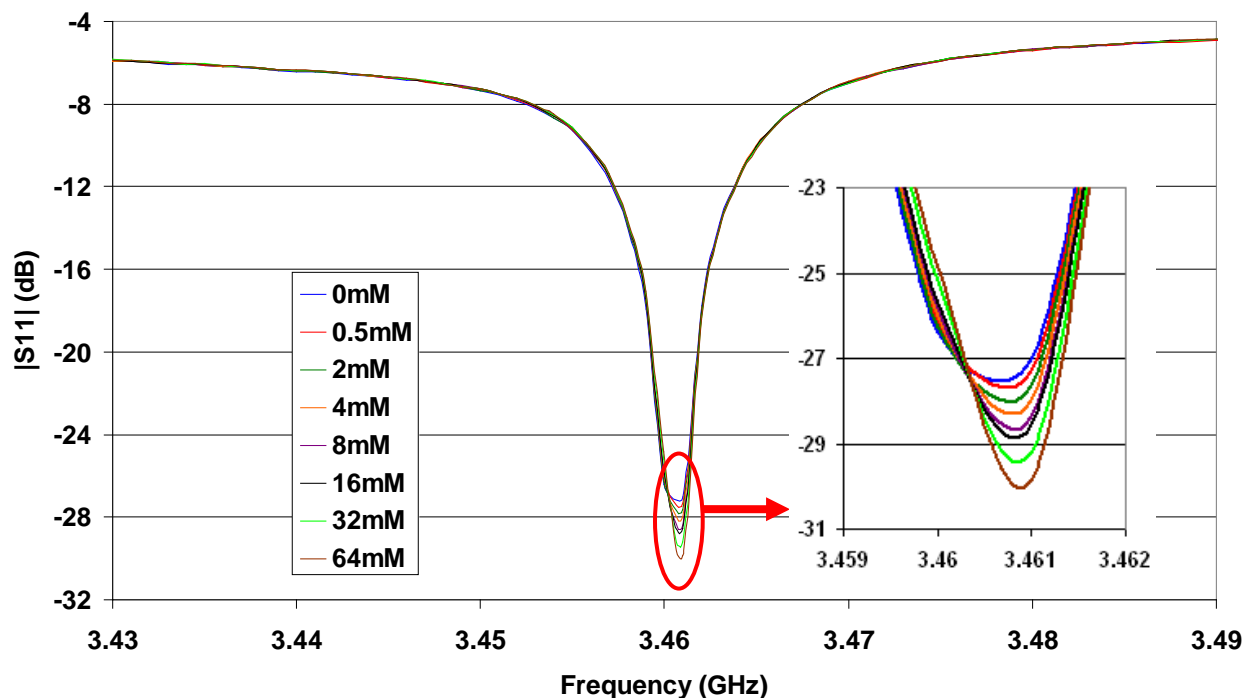


Figure 7: The $|S_{11}|$ spectrum for 10ml lactate in distilled water using the large volume cavity; the frequency range of interest here is 3.43-3.49GHz, with resonance occurring at 3.461GHz.

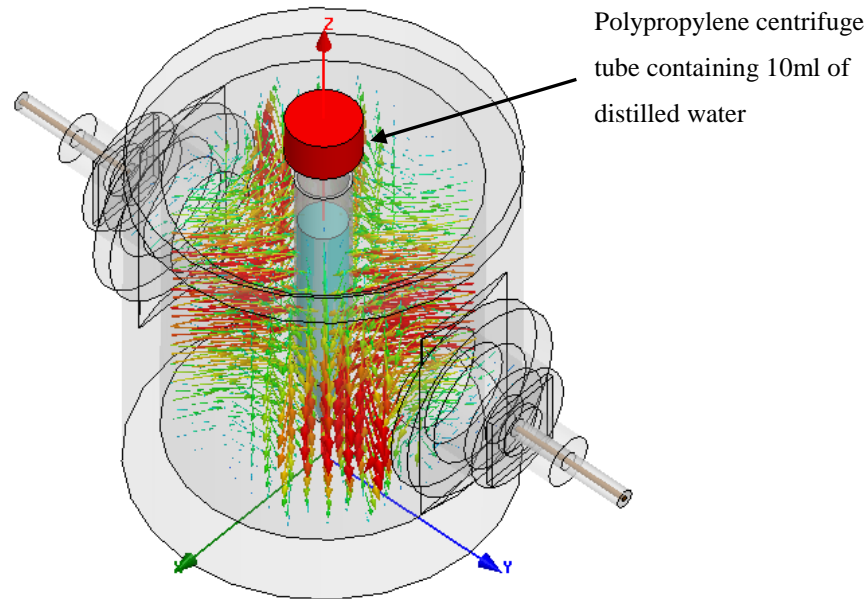


Figure 8: TE₂₁₁ mode for large volume cavity, simulated with a water sample at the centre.

b. Small volume cavity

Figure 9 shows the $|S_{11}|$ spectrum for the small volume cavity, which was used to analyse the 0.5ml samples. From this spectrum result, one can see the level of amplitude increase as the concentration of the lactate increases from 0mM (distilled water) to 64mM at around 1.625GHz. It is difficult to give a definitive figure here since the frequency shifts with concentration in a non-linear way. These results do however show a linear trend for signal amplitude when the concentration of lactate varies in water. The lowest amplitude (-13.62dB) occurs when analysing the 0mM sample, whilst the highest amplitude (-9.33dB) occurs for 64mM sample. Across the complete sample set, the amplitude increases with lactate concentration at a rate of 0.067dB/mM.

As described with the large volume cavity, it is possible to determine the mode at which this response occurs by matching the measured resonance to that of the simulated model in HFSS, which now includes the sample container with 0.5ml of water. Figure 10 shows the simulated model for the small volume cavity at 1.624GHz, where the TM₀₁₀ mode dominates. In this case the error between the measured and simulated resonance is 1MHz, which is better than before and therefore acceptable.

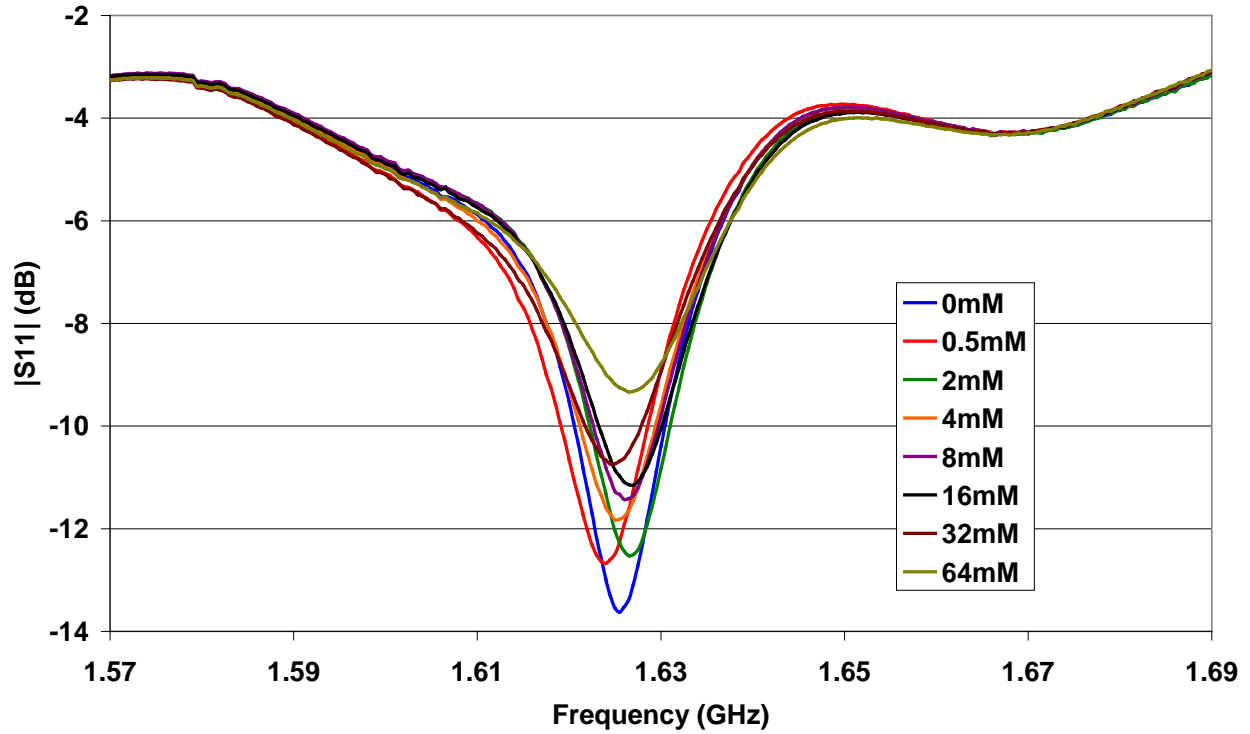


Figure 9: The $|S_{11}|$ spectrum for 0.5ml lactate in distilled water by using the small volume cavity; the frequency range of interest here is 1.57-1.69GHz, with resonance occurring at 1.625GHz.

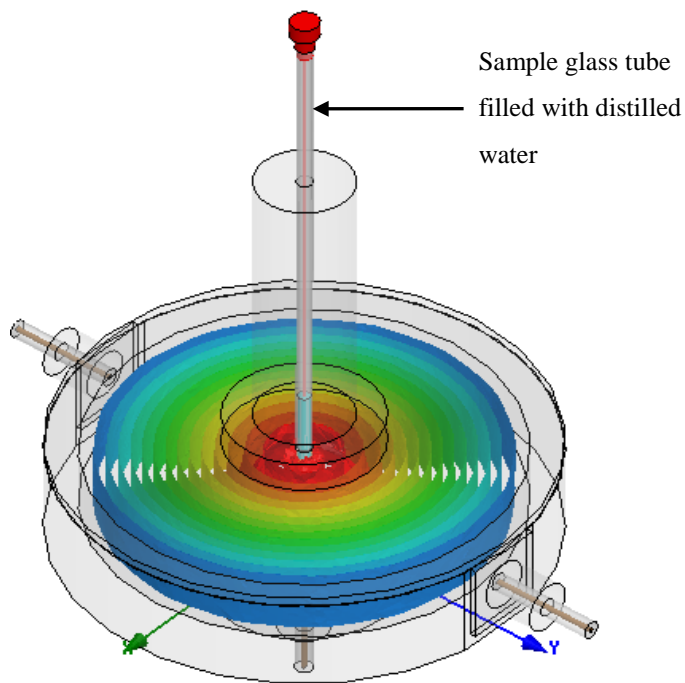


Figure 10: TM_{010} mode for small volume cavity with sample glass tube at centre.

V. CONCLUSIONS

Results from this work, which considers varying lactate concentrations in water, show that a linear response is possible when using microwave cavity sensors. The work has demonstrated the use of two different sensors, one which accommodates large volumes (10ml) and one which accommodates smaller volumes (0.5ml). Signal amplitude decreases by 0.04dB/mM when using large volume cavity, and increases by 0.067dB/mM when using the small volume cavity. Whilst these results alone would suggest that the small volume cavity provides better sensitivity *and* smaller sample size, it does not provide a stable resonant frequency and therefore may prove to be less precise than the large volume cavity.

Despite this finding, the work thus far suggests that microwave spectroscopy holds significant potential for the detection of issues during surgery when presented with CSF samples. Future elements of the work will include expanding into other clinically useful indicators in CSF such as albumin, glucose, lactate dehydrogenase (LDH), and consider the ability to rapidly scan biological fluids and tissues *ex-vivo* for cancer cells or tumour architecture. Ultimately however, the ability to gain useful information from microwave scans of tissues *in-vivo* could serve to avoid a range of invasive diagnostic procedures.

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